学位論文全文に代わる要約 Extended Summary in Lieu of Dissertation

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学位論文題目: Title of Dissertation Evaluation of Allelopathic Activity and Identification of Allelopathic Substances in Myanmar Medicinal Plants (ミャンマー在来薬用植物のアレロパシー活性とアレロパシー物質の同定)

学位論文要約: Dissertation Summary

Weed management is crucial for crop production, as they constantly compete with crop plants, causing significant crop productivity losses. In modern agricultural practice, herbicides have played a central role in agricultural intensification that has resulted in progressive increases in crop yields worldwide (Gianessi, 2013), but in many cases, this increase in crop production has come at a cost of environmental degradation (Barbash et al., 2001; Davis et al., 2014). Herbicide-resistant weeds have also been reported for 96 species of crops in 71 countries (Heap, 2022). Besides, several countries have recently banned different herbicides in response to health and environmental issues posed by their widespread use (Adhikari et al., 2019). Many herbicide-resistant weed species occur in the major field crops grown worldwide (e.g., canola, cotton, maize, rice, sorghum, soybeans, winter-, spring- and durum-wheat), as well as pulse crops and sown pastures (Heap, 2014). Furthermore, the concerning negative effects of herbicides are increasing (Gandhi and Snedeker, 1999), leading to the development of alternative weed management technologies. 1999). So, many efforts have been deployed in designing alternative weed management technologies. Alternative weed management technologies based on natural products such as the use of allelopathy phenomenon, cover crops and living mulches, competitive crop cultivars, and suitable nutrient management can be proposed as low-cost, effective, and eco-friendly practices for sustainable weed management in cropping systems (Hasan et al., 2021). These practices can promote soil health and biodiversity, leading to long-term benefits for the environment and crop production.

Allelopathy refers to the direct or indirect effect of plants upon neighboring plants or their associated microflora or microfauna by the production of allelochemicals that interfere with the growth of the plant (IAS, 2018). The utilization of allelopathic properties in agricultural management has been extensively studied, and several research studies suggest that allelopathy tends to develop as an alternative strategy for weed management (An et al., 1998; Macías et al., 1999; Khanh et al., 2005; Sodaeizadeh and

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Zahra, 2012). Allelochemicals are produced by plants as secondary metabolites or by microbes through decomposition, and their mode of action, uptake, and effectiveness is still unclear (Rice, 1984). These allelochemicals can suppress germination, growth, and establishment of surrounding plants or modify soil properties in the rhizosphere by influencing the microbial community (Weir et al., 2004; Zhou et al., 2013). Many plant species have been listed worldwide for their allelopathic effects, with some having great potential for further research (Duke et al., 2002). Researchers worldwide are increasingly interested in medicinal plants for their potential therapeutic properties (Azizi and Fuji, 2006; Han et al., 2008; Li et al., 2009). This interest is driven by two main reasons: the easier screening process of phytotoxic plants from medicinal plants and the possibility of having more bioactive compounds in medicinal plants (Islam and Kato-Noguchi, 2014). Several researchers for example, Anjum et al. (2010), Laosinwattana et al. (2012), Khan et al. (2013), Algandaby and El-Darier (2016), Qasem (2017), Kyaw et al. (2022) and Moh et al. (2023) worked with some medicinal plants species and observed that most of the species possessed strong allelopathic properties and inhibited the growth of test plant species with different inhibition values.

This study has focused on the evaluation of allelopathic activity and the identification of allelopathic substances in Myanmar medicinal plants. Plumbago rosea Linn. is a perennial flowering plant belonging to the family of Plumbaginaceae, that grows well in tropical climates, tropical Africa, tropical Asia, and the Pacific region. Polygonum chinense Linn. is a perennial herb plant belonging to the family of Polygonaceae, that is reported from Myanmar, Thailand, China, and Northeast India and is endemic to Southeast Asia (Paul and Chowdhury, 2016). Acmella uliginosa (Sw.) Cass is an annual herb belonging to the family of Asteraceae, are traditional medicinal plants that is extensively distributed in tropic and subtropic regions mainly in Brazil, Africa, Indonesia, West Indies, and Malaysia (Pandey et al., 2007). The pharmacological properties of these three plants are well documented (Dorni et al., 2006, Lai et al., 2012, Sana et al., 2014) and they constitute many active biochemicals (Devi et al., 1998, Manasa et al., 2016, Gupta et al., 2012). Because of its different bioactivities, it is believed that it may also possess allelopathic activity. However, there has been no research, report, or information on its allelopathic properties yet. Therefore, the allelopathy potentials of these three medicinal plants, Plumbago rosea Linn., Polygonum chinense Linn., and Acmella uliginosa (Sw.) Cass were studied.

Stems of *Plumbago rosea*, plants of *Polygonum chinense*, and plants of *Acmella uliginosa* were collected from the area of Mandalay Division, Myanmar in July-August 2020. The plants along with the roots were removed from the adhering soils by thoroughly washing through running water, then spread out and allowed to dry in the shade with

adequate airflow. The dried materials were ground into a fine powder and kept in the vacuum-sealed package at 4°C for further extraction. The dried powder of *P. rosea, P. chinense*, and *A. uliginosa* was separately extracted with 70% (v/v) aqueous methanol by soaking it in a closed container for 48 h. The extract was filtered using a vacuum pump and the residue of plant material was extracted again with 2.5 L of cold methanol for 24 h. The two filtrates were combined and evaporated with a rotary evaporator at 40°C until dryness. The species for determination of biological activity were composed of both dicotyledonous and monocotyledonous plant species. The representative test plant species consisted of cress (*Lepidium sativum* L.), lettuce (*Lactuca sativa* L.), alfalfa (*Medicago sativa* L.), barnyard grass (*Echinochloa crus-gallis* (L.) P. Beauv), Italian ryegrass (*Lolium multiflorum* Lum.), and timothy (*Phleum pretense* L.).

The results showed that the growth inhibitory effect on aqueous methanol stems extracts of Plumbago rosea, plant extracts of Polygonum chinense and plant extracts of Acmella uliginosa were evaluated by six test plant species of timothy, barnyard grass, Italian ryegrass, cress, lettuce, and alfalfa. The extract concentration obtained from 30 mg dry weight equivalent extracts/mL of all the tested plants was inhibited by more than 50 % except the shoot growth of the barnyard grass. At the highest concentration of 300 mg dry weight equivalent extracts/mL, the shoot and root growth of all the tested plants were completely inhibited (100%) except the shoot of barnyard grass when treated with P. rosea and P. chinense (> 94%) and the root growth of alfalfa when treated with A. uliginosa (>94%). The inhibitory effect is augmented with increasing concentrations of methanol extracts. The effectiveness of the extract depended on the extract concentration and species. Besides, differences in the growth inhibition resulting from the variation of extract concentrations and specificity of the test plants were also reported by Parvez et al., 2004, Shunjie et al., 2008, Moosavi et al., 2011; Moh et al., 2023. According to the I₅₀ values of P. rosea, P. chinense, and A. uliginosa, the shoot length ranged from 0.87 to 33.5, 3.5 to 35.09, and 2.9 to 56.79 mg DW equivalent extract/mL, respectively, and the root length ranged from 1.13 to 11.68, 2.54 to 11.7, and 2.47 to 21.56 mg DW equivalent extract/mL, respectively. Based on the I_{50} values of aqueous methanol extracts of *P. rosea*, P. chinense, and A. uliginosa showed different responses on the shoot and root growth of six test plant species in a species-specific manner. Other researchers have also documented the sensitivity of different test plant species to the varies of extracts (Zaman et al., 2020; Krumsri et al., 2021; Kyaw et al., 2022). For almost all the root lengths of the test plant species, the extract had greater inhibitory effects on root growth than their shoot growth. A similar result was reported by Arowosegbe and Afolayan (2012) that the extracts of Aloe ferox had greater inhibitory effects on roots than shoots of some test plants. This may be because the root is the first organ that exposes to the extracts and absorbs allelochemicals

(Salam and Kato-Noguchi, 2010). The growth inhibitory potentials of the extracts from these extracts suggest that these medicinal plants have allelopathic effects and might possess allelopathic compounds. Therefore, these medicinal plants might be used as suitable candidates for the isolation and characterization of potent allelopathic compounds.

Afterward, all plant extracts were separately adjusted to pH 7.0 with 1 M phosphate buffer and partitioned four times with an equal volume of ethyl acetate (EtOAc). The EtOAc and aqueous fractions determined the inhibitory activity on the growth of cress. The EtOAc fractions were carried out to isolate and purify the growth inhibitory substances through several chromatographic fractionations: silica gel column, Sephadex LH-20, reverse-phase C₁₈ cartridge, and reverse-phase HPLC. In every purification step of HPLC analysis, the wavelength of the UV/VIS detector was 220 nm. The chemical structures of isolated substances were determined using APCIMS, HRESIMS, ¹H- and ¹³C NMR. The biological effects of all isolated substances were determined by the seedlings' growth of the cress. All treatments were prepared with a completely randomized complete design.

Two growth inhibitory substances were observed from *Plumbago rosea* stem identified 7,3',4'-tri-O-methyl extracts and as dihydroquercetin, and 7,4',5'-tri-O-methylampelopsin. The two compounds are structural derivatives of flavonoids, which are the largest group of naturally occurring phenols (Hussein and El-Anssary, 2018). Many flavonoids of similar structures are known to be effective radical scavengers (antioxidants) (Tuckmantel et al., 1999). Ampelopsin has the ability to inhibit cell proliferation, migration, and invasion in breast and prostate cancer (Ni et al., 2012, Zhou et al. 2014). Yang et al. (2020) reported for the first time that ampelopsin can simultaneously inhibit the proliferation of melanoma tumor cells. The two compounds inhibited significantly the shoot and root growth of the cress at the concentration of 0.3 mM. The I₅₀ values of 7,4',5'-tri-O-methylampelopsin, and 7,3',4'-tri-O-methyl dihydroquercetin on the root and shoot growth of cress were 0.07 to 0.21 mM and 0.24 to 0.59 mM, respectively. Comparing the I_{50} values of the compounds indicate that 7,4',5'-tri-*O*-methylampelopsin possesses greater allelopathic activity than 7,3',4'-tri-O-methyl dihydroquercetin. This finding may be due to the 3'-OH group in 7,4',5'-tri-*O*-methylampelopsin ring Β. Some researchers have reported that 3,5,7,3',4',5'-hexa-hydroxyflavone is a stronger antioxidant than quercetin, which has been attributed to the 5'-OH group that allows further stabilization of the myricetin-derived semi-quinone radical (Oyama et al., 1994; Gordon and Roedig-Penmanm, 1998). Several studies documented that the toxicity of phytochemicals is regulated by their chemical structure, particularly the number and position of various functional groups substituted in the benzene ring (Cueva et al., 2010; Sanchez-Maldonado et al., 2011). However, limited

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information concerning the allelopathic activities of 7,3',4'-tri-O-methyl dihydroquercetin, and 7,4',5'-tri-O-methylampelopsin; this study is the first to report on the allelopathic activities of 7,3',4'-tri-O-methyl dihydroquercetin and 7,4',5'-tri-O-methylampelopsin. The present results suggest that the allelopathic effect of *P. rosea* stem and their identified compounds exert could potentially be used as a natural source of herbicide for controlling the weeds.

Four active inhibitory substances were isolated from Polygonum chinense plant extracts and identified as 1. dehydrovomifoliol, 2. (-)-3-hydroxy- β -ionone, 3. (-)-3-hydroxy-7,8-dihydro- β -ionone and 4. loliolide. All of the four compounds are norisoprenoid aglycons, plant constituents, which are generally considered to be derived from carotenoids by oxidative degradation (Enzell, 1985). Dehydrovomifoliol and loliolide are reported carotenoid metabolites (Pan et al. 2009), and their antimicrobial, antiproliferative, anti-algal, antioxidant, and cytotoxic properties have been explored (Ragasa et al., 2005; Lu et al., 2011), they can also be synthesized from C₉-hydroxy ketone and C₁₁-aldehyde, respectively (Mayer, 1979). Ren et al. (2009) reported that dehydrovomifoliol cytotoxic effects has against human cancer cells. (-)-3-hydroxy- β -ionone has been documented as a bound constituent of several fruit tissues, such as apple (Enzell 1985), grape (Schwab and Schreier, 1988), and papaya (Strauss et al., 1987) and it accumulates in the seedlings of bean varieties through irradiation by light, causing light-induced growth inhibition of bean seedlings (Schwab et al., 1989, Kato-Noguchi, 1992). (-)-3-Hydroxy-β-ionone has also been isolated and identified from various plants, and its growth inhibition potential against a number of species is well reported (Fujimori et al. 1974; Kato-Noguchi et al. 2010; Masum et al. 2018; Ida et al. 2020; Hossen et al., 2022). Aloum et al. (2020) also reported that (-)-3-hydroxy- β -ionone retards the colony formation, proliferation, and cell migration of human squamous cell carcinoma. (-)-3-Hydroxy-7,8-dihydro-\beta-ionone has already been described as a conjugate in the aqueous extract of Epimedium grandiflorum var. thunbergianum (Miyase et al., 1988). Loliolide was first reported from Lolium perenne L. by Hodges and Porte (1964) and has diverse biological activities. (Yang et al. 2011; Okada et al. 1994, Chung et al. 2016; Okunade and Wiemer, 1985). Although dehydrovomifoliol, (-)-3-hydroxy-β-ionone, (-)-3-hydroxy-7,8-dihydro-β-ionone and loliolide have been documented in many plants, there is no report from Polygonum chinense yet. Therefore, this study is the first to document the presence of these four compounds in Polygonum chinense extracts and their potentially allelopathic activity. At the concentration of 0.03 mM, (-)-3-hydroxy-β-ionone, (-)-3-hydroxy-7,8-dihydro-β-ionone, and lolilide have a significant inhibitory effect against the shoot growth of cress whereas dehydrovomifoliol significantly inhibited at the concentration of 10 mM. The root growths of cress were significantly inhibited by Compounds 1-4 at the concentration of 1 mM, 0.3 mM, 0.3mM, and 0.1 mM, respectively. Compounds (1-3) have 13 carbon atoms and similar structures with different functional groups. Dehydrovomifoliol had two oxo groups at C-1 and C-9 whilst the Compounds (2-3) had one oxo group. In (-)-3-hydroxy- β -ionone, the C-7 and C-8 positions are linked with the carbon-carbon double bond (olefins carbon) functional group, while (-)-3-hydroxy-7,8-dihydro-β-ionone is linked with a carbon-carbon single bond in the functional group. Loliolide consists of 11 carbon atoms arranged in a benzene ring with a hydroxyl group combined with a five-membered lactone. The I_{50} values of compounds 1-4 for the shoot and root growth of cress were 2.58 and 1.7 mM, 0.05 and 0.07 mM, 0.42 and 1.29 mM, 0.25 and 0.47 mM, respectively, and for compound 2 were 0.42 and 1.29 mM, respectively. Whilst comparing the I_{50} values of the four compounds, (-)-3-hydroxy- β -ionone had the strongest inhibition effect followed by loliolide, (-)-3-hydroxy-7,8-dihydro- β -ionone and dehydrovomifoliol at lower concentrations. Their different presence of the olefin carbon functional group, hydroxy group, and oxo group may be the reason for their different allelopathic activities. Allelopathic compounds vary in mode of action, uptake, and effectiveness (Weston and Duke, 2003; Rice, 2012). These findings indicate that these four substances possess potentially allelopathic properties and may make an important contribution to the allelopathic substances of P. chinense. Hence, P. chinense might be useful in weed management through the application of its extracts, the inclusion of its residues or different parts as mulch, and the application of its active substances as natural product-based agriculture to reduce synthetic chemical herbicides usage, and also to attain sustainable crop production for pollutant-free green environments.

The bioassay-directed fractionations of *Acmella uliginosa* whole plant extracts indicated that the extract contains some potential inhibitory substance, and the active substances were isolated with a strong growth inhibitory activity. The inhibitory substances of *A. uliginosa* whole plant extracts will further be identified as the chemical structures which may provide the chemical basis for the development of bioherbicides and may play an important role in controlling weed growth as alternative weed management. So far, there is no report on the identification of inhibitory substances against the growth of the test plants and these active peaks may be responsible for the allelopathic activity of *A. uliginosa*.

In conclusion, the present study revealed that the aqueous methanol extracts of stems of *Plumbago rosea*, the whole plant of *Polygonum chinense*, and the whole plant of *Acmella uliginosa* growth inhibitory activity against different test plants of three monocotyledonous and three dicotyledonous species, indicating these plant materials may contain substances with allelopathic potential. Consequently, six potential allelopathic substances were isolated and identified as; 7,3',4'-tri-*O*-methyl dihydroquercetin, and

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7,4',5'-tri-O-methylampelopsin from the stem extracts of *Plumbago rosea* and dehydrovomifoliol, (-)-3-hydroxy- β -ionone, 3-hydroxy-7,8-dihydro- β -ionone and loliolide from the whole plant extracts of *Polygonum chinense*. It is noted that all the isolated substances also showed significant inhibitory activity on the test plants in a concentrationand species- specific manner. Therefore, it can be summarized that these substances may be involved in the allelopathic activity of the respective plant materials from where they were isolated. Finally, the application of those isolated substances and/or the plant parts might be utilized as a soil additive or soil amendment to control weeds in an eco-friendly way and may be released into the soil by the decomposition of its plant residues, and they might act as allelopathic substances. However, more research is needed to examine the mechanisms underlying the allelopathy of its plants, the role of allelochemicals of different functional groups, and the long-term effects of allelopathy in soil residues.

References

- Adhikari, S.P., Yang, H. and Kim, H. (2019) Learning semantic graphics using convolutional encoder-decoder network for autonomous weeding in paddy. Front. Plant Sci. 10: 1404.
- Algandaby, M.M. and El-Darier, S. (2016) Management of the Noxious Weed; *Medicago polymorpha* L. via Allelopathy of Some Medicinal Plants from Taif Region, Saudi Arabia. Saudi J. Biol. Sci. 25, 1339-1347.
- Aloum, L., Alefishat, E., Adem, A. and Petroianu, G. (2020) Ionone Is More than a Violet's Fragrance: A Review. *Molecules*. 25, 5822; DOI:10.3390/molecules25245822
- An, M., Pratley, J. and Haig, T. (1998) Allelopathy: from concept to reality. In Proceedings of the 9th Australian Agronomy Conference, Wagga Wagga, Australia. 563–566. Available online: http://www.regional.org.au/au/asa/1998/6/314an.htm (October 20, 2016).
- Anjum, A., Hussain, U., Yousaf, Z., Khan, F. and Umer, A. (2010) Evaluation of Allelopathic Action of Some Selected Medicinal Plant on Lettuce Seeds by Using Sandwich Method. J. Medicinal Plants, 4, 536-541.
- Arowosegbe, S. and Afolayan, A.J. (2012) Assessment of allelopathic properties of *Aloe ferox* Mill. on turnip, beetroot, and carrot. Biol. Res. 45: 363–368.
- Azizi, M. and Fujii, Y. (2006) Allelopathic effect of some medicinal plant substances on seed germination of Amaranthus retroflexus and Portulaca oleraceae. Acta Hortic. 699:61-68.
- Barbash, J.E., Thelin, G.P., Kolpin, D.W. and Gilliom, R.J. (2001) Major herbicides in groundwater: results from the national water-quality assessment. J. Environ. Qual.

30, 831-845.

- Chung, C.-Y., Liu, C.-H., Burnouf, T., et al. (2016) Activity-based and fraction-guided analysis of *Phyllanthus urinaria* identifies loliolide as a potent inhibitor of hepatitis C virus entry. *Antiviral Res.* 130, 58-68.
- Cueva, C., Moreno-Arribas, M.V., Martinez-Alvarez, P.J., Bills, G., Vicente, M.F., Basilio,
 A., Lopez Rivas, C., Requena, T., Rodríguez, J.M. and Bartolomé, B. (2010)
 Antimicrobial activity of phenolic acids against commensal, probiotic and pathogenic bacteria. *Res. Microbiol.* 16, 372–382.
- Davis, A.M., Lewis, S.E., Brodie, J.E. and Benson, A. (2014) The potential benefits of herbicide regulation: a cautionary note for the Great Barrier Reef catchment area. *Sci. Total Environ.* 490, 81–92.
- Devi, P.U., Rao, B.S. and Solomon, F.E. (1998) Effect of plumbagin on the radiation-induced cytogenetic and cell cycle changes in mouse Ehrlich ascites carcinoma *in vivo*. *Indian J Exp Biol*. 36, 891–895.
- Dorni, A.I.C., Vidyalakshmi, K.S., Hannah, R.V., Rajamanickam, G.V. and Dubey, G.P. (2006) Anti-inflammatory activity of *Plumbago capensis*. *Pharmacognosy Mag.* 2, 239–243.
- Duke, S.O., Dayan, F.E., Romagni, J.G. and Rimando, A.M. (2002) Natural products as sources of herbicides: current status and future trends. Weed Res. 40 (1), 99–111. DOI: 10.1046/j.1365-3180.2000.00161.x
- Enzell, C. (1985) Biodegradation of carotenoids-an important route to aroma components. *Pure Appl. Chem.* 57, 693–700.
- Fujimori, T., Kasuga, R., Noguchi, M. and Kaneko, H. (1974) Isolation of R-(-)-3-hydroxy-β-ionone from burley tobacco. Agric Biol Chem. 38:891–892.
- Gandhi, R. and Snedeker, S.M. (1999) Critical evaluation of dichlorvos breast cancer risk.
 Critical evaluation #7. Program on breast cancer and environmental risk factors in New York State (BCERF), Cornell UniversityGilani, S.A., Fujii, Y., Shinwari, Z.K., Adnan, M., Kikuchi, A. and Watanabe, K.N. (2010) Phytotoxic Studies of Medicinal Plant Species of Pakistan. Pakistan *Journal of Botany*. 42, 987-996.
- Gordon, M.H. and Roedig-Penmanm, A. (1998) Antioxidant activity of quercetin and myricetin in liposomes. *Chem Phys Lipids*. 97, 79–85. DOI:10.1016/S0009-3084(98) 00098-X
- Gupta, N., Patel A.R., and Ravindra R.P. (2012) Int. J. Pharm. Bio Sci., 3 (4), 161–170.
- Han, C.M., Pan, K.W., Wu, N., Wang, J.C., and Li, W. (2008) Allelopathic Effect of Ginger on Seed Germination and Seedling Growth of Soybean and Chive. *Scientia Horticulture*. 116, 330-336. <u>https://doi.org/10.1016/j.scienta.2008.01.005</u>

Hasan, M., Ahmad-Hamdani, M.S., Rosli, A.M. and Hamdan, H. (2021) Bioherbicides: An

Eco-Friendly Tool for Sustainable Weed Management. *Plants.* 10, 1212. https://doi.org/10.3390/ plants10061212 Academic Editor: Fab

- Heap, I. (2014) Global perspective of herbicide-resistant weeds. *Pest Manag. Sci.* 70, 1306–1315.
- Heap, I. (2022) The International Survey of Herbicide Resistant Weeds. (www. weedscience.com).
- Hodges, R., Porte, A.L. (1964) The structure of loliolide: A terpene from *Lolium perenne*. *Tetrahedron*. 20: 1463–1467.
- Hossen, K., Das, K.R., Asato, Y., Teruya, T. and Kato-Noguchi, H. (2022) Allelopathic activity and characterization of allelopathic substances from *Elaeocarpus floribundus* Blume leaves for the development of bioherbicides. Agronomy. 12, 57. https://doi.org/10.3390/ agronomy12010057
- Hussein, R.A. and El-Anssary, A.A. (2018) Plants secondary metabolites: The key drivers of the pharmacological actions of medicinal plants. In: Builders, P. (Ed.) Herbal Medicine. Intech Open. Chapter 2.
- IAS. International Allelopathy Society. (2018) http://allelopathy-society.osupytheas.fr/about/
- Ida, N., Iwasaki, A., Teruya, T., Suenaga, K. and Kato-Noguchi, H. (2020) Tree fern *Cyathea lepifera* may survive by its phytotoxic property. *Plants.* 9, 46.
- Islam, A.K.M.M and Kato-Noguchi, H. (2014) Phytotoxic Activity of Ocimum tenuiflorum Extracts on Germination and Seedling Growth of Different Plant Species. Sci. World J. Article ID: 676242.
- Kato-Noguchi, H. (1992) An endogenous growth inhibitor, 3-hydroxy- β -ionone. I. Its role in light-induced growth inhibition of hypocotyls of *Phaseolus vulgaris*. *Physiol. Plant.* 86: 583–586.
- Kato-Noguchi, H., Ino, T. and Kujime, H. (2010) The relation between growth inhibition and secretion level of momilactone B from rice root. J. Plant Interact. 5: 87–90.
- Khan, M.S.I., Islam, A.K.M.M. and Kato-Noguchi, H. (2013) Evaluation of Allelopathic Activity of Three Mango (*Mangifera indica*) Cultivars. Asian J. Plant Sci. 12, 252-261. <u>https://doi.org/10.3923/ajps.2013.252.261</u>
- Khanh, T.D., Hong, N.H., Xuan, T.D. and Chung, I.M. (2005) Paddy Weed Control by Medicinal and Leguminous Plants from Southeast Asia. Crop Protection. 24, 421-431. <u>https://doi.org/10</u>. 1016/j.cropro.2004.09.020
- Krumsri, R., Ozaki, K., Teruya, T. and Kato-Noguchi H. (2021) Isolation and identification of two potent phytotoxic substances from *Afzelia xylocarpa* for controlling weeds. *Appl. Sci.* 11: 3542.
- Kyaw, E.H., Iwasaki, A., Suenaga, K. and Kato-Noguchi, H. (2022) Assessment of the

Phytotoxic Potential of *Dregea volubilis* (L.f.) Benth. ex Hook. f. and Identification of its Phytotoxic Substances for Weed Control. *Agriculture* 12, 1826. https://doi.org/10.3390/agriculture12111826

- Lai, S.M., Sudhahar, D. and Anandarajagopal, K. (2012) Evaluation of Antibacterial and Antifungal activities of *Persicaria chinensis* Leaves. Int. J. Pharm. Sci. Res. 3, 2825–2830.
- Laosinwattana, C., Teerarak, M. and Charoenying, P. (2012) Effects of Aglaia odorata Granules on the Seedling Growth of Major Maize Weeds and the Influence of Soil Type on the Granule Residue's Efficacy. Weed Biol. Manag. 12,117-122. https://doi.org/10.1111/j.1445-6664.2012.00444.x
- Li, H., Pan, K., Liu, Q. and Wang, J. (2009) Effect of Enhanced Ultraviolet-B on Allelopathic Potential of Zanthoxylum bungeanum. Sci. Hortic. (Amsterdam). 119, 310-314. https://doi.org/10.1016/j. scienta.2008.08.010
- Lu, H., Xie, H., Gong, Y., Wang, Q. and Yang, Y. (2011) Secondary metabolites from the seaweed Gracilaria lemaneiformis and their allelopathic effects on Skeletonema costatum. Biochem. Syst. Ecol. 39, 397–400.
- Macías, F.A., Oliva, R.M., Varela, R.M., Torres, A. and Molinillo, J.M. (1999) Allelochemicals from sunflower leaves cv. *Peredovick*. *Phytochemistry*. 52, 613–621.
- Manasa, K.S., Kuppast, I.J., Kishan, K.M.A. and Akshara, K. (2016) A review on *Polygonum chinensis. Res. j. pahrmacol. pharmacodyn.* 8, 185–188. DOI: 10.5958/23215836.2016.00034.3
- Masum, S.M., Hossain, M.A., Akamine, H., Sakagami, J.I., Ishii, T., Gima, S., Kensaku, T. and Bhowmik, P.C. (2018) Isolation and characterization of allelopathic compounds from the indigenous rice variety 'Boterswar' and their biological activity against *Echinochloa crus-galli* L. *Allelopath. J.* 43, 31–42.
- Mayer, H. (1979) Synthesis of optically active carotenoids and related compounds. *Pure* Appl Chem. 51: 535–564.
- Miyase, T.; Ueno, A.; Takizawa, N.; Kobayashi, H.; Oguchi, H. Studies on the glycosides of *Epimedium grandiflorum* var *thunbergianum* miq. Nakai. *Chem. Pharm. Bull.* 1988, 36, 2475–2484.
- Moh, S.M., Kurisawa, N., Suenaga, K. and Kato-Noguchi, H. (2023) Allelopathic Potential of *Marsdenia tenacissima* (Roxb.) Moon against Four Test Plants and the Biological Activity of Its Allelopathic Novel Compound, 8-Dehydroxy-11β-OAcetyl-12β-O-Tigloyl-17β-Marsdenin. *Plants.* 12, 1663. <u>https://doi.org/10.3390/plants12081663</u>
- Moosavi, A., Afshari, R.T., Asadi, A. and Gharineh, M. H. (2011) Allelopathic effects of aqueous extract of leaf stem and root of *Sorghum bicolor* on seed germination and seedling growth of *Vigna radiata* L. *Not. Sci. Biol.* 3: 114–118.

- Ni F, Gong Y, Li L, Abdolmaleky HM, Zhou J-R. (2012) Flavonoid ampelopsin inhibits the growth and metastasis of prostate cancer *in vitro* and in mice. *PLoS One*. 7(6):e38802.
- Okada, N., Shirata, K., Niwano, M., et al. (1994) Immunosuppressive activity of a monoterpene from *Eucommia ulmoides*. *Phytochemistry*. 37(1), 281-282 (1994).
- Okunade, A.L., and Wiemer, D.F. (1985) (-)-Loliolide, an ant-repellent compound from Xanthoxyllum setulosum. J. Nat. Prod. 48(3), 472-473.
- Oyama, Y., Fuchs, P.A., Katayama, N. and Noda, K. (1994) Myricetin and quercetin, the flavonoid constituents of *Ginkgo biloba* extract, greatly reduce oxidative metabolism in both resting and Ca⁽²⁺⁾⁻ loaded brain neurons. *Brain Res.* 635, 125–129. DOI:10.1016/0006- 8993(94)91431-1
- Pan, L., Sinden, M.R., Kennedy, A.H., Chai, H., Watson, L.E., Graham, T.L., Kinghorn,
 A.D. (2009) Bioactive constituents of *Helianthus tuberosus* (Jerusalem artichoke). *Phytochem Lett.* 2: 15–18.
- Pandey, V., Agrawal, V., Raghavendra, K. and Dash, A.P. (2007) Strong larvicidal activity of three species of Spilanthes (Akarkara) against malaria (Anopheles stephensi Liston, Anopheles culicifacies, species C) and filaria vector (Culex quinquefasciatus Say). Parasitol. Res. 102(1), 171-174.
- Parvez, S.S., Parvez, M.M., Fujii, Y. and Gemma, H. (2004) Differential allelopathic expression of bark and seed of *Tamarindus indica* L. *Plant Growth Regulation*. 42: 245–252.
- Paul, P. and Chowdhury, M. (2016) Persicaria chinensis (Linnaeus) H. Gross var. hispida (Hooker f.) Kantachot (Polygonaceae): a new distributional record for West Bengal, India. Asian J. Biol. Life Sci. 5, 260-262.
- Qasem, J.R. (2017) A Survey on the Phytotoxicity of Common Weeds, Wild Grown Species and Medicinal Plants on Wheat. *Allelopathy Journal*. 42, 179-194. https://doi.org/10.26651/allelo.j./2017-42-2-1115
- Ragasa, C.Y., De, Luna, R.D. and Hofilena, J.G. (2005) Antimicrobial terpenoids from *Pterocarpus indicus. Nat. Prod. Res.* 19, 305–309.
- Ren, Y., Shen, L., Zhang, D. and Dai, S. (2009) Two New sesquiterpenoids from *Solanum lyratum* with cytotoxic activities. *Chem. Pharm. Bull.* 57(4): 408–410.
- Rice, E. L. (1984) Allelopathy. 2nd Edn, Orlando, FL: Academic press.
- Rice, E.L. (2012) Allelopathy (New York, NY: Academic Press). 104–125.
- Salam, M.A. and Kato-Noguchi, H. (2010) Evaluation of allelopathic potential of neem (Azadirachta indica. A. Juss) against seed germination and seedling growth of different test plant species. International Journal of Sustainable Agriculture. 2: 20-25.

- Sana, H., Rani, A.S. and Sulakshana, G. (2014) Determination of antioxidant potential in *Spilanthes acmella* using DPPH assay. *Int. J. Curr. Microbiol. Appl. Sci.* 3, 219-213.
- Sanchez-Maldonado, A.F., Schieber, A. and Ganzle, M.G. (2011) Structure-function relationships of the antibacterial activity of phenolic acids and their metabolism by lactic acid bacteria. J. Appl. Microbiol. 111, 1176–1184.
- Schwab, W., and Schreier, P. (1988) Simultaneous enzyme catalysis extraction: A versatile technique for the study of flavor precursors. J. Agric. Food Chem. 36, 1238–1242.
- Schwab, W., Mahr, C. and Schreier, P. (1989) Studies on the enzymic hydrolysis of bound aroma components from *Carica papaya* fruit. J. Agric. Food Chem. 37, 1009–1012.
- Shunjie, Z., Ma, F., Yubo, W. and Zhen, S. (2008) Study on allelopathy of soybean root exudates. J. Northeast Agric. Univ. 10: 83-91.
- Sodaeizadeh, H. and Zahra, H. (2012) Allelopathy an environmentally friendly method for weed control. International conference on applied life sciences., Turkey. 387–392.
- Strauss, C.R., Gooley, P.R., Wilson, B., and Williams, P.J. (1987) Application of droplet countercurrent chromatography for the analysis of conjugated forms of terpenoids, phenols, and other constituents of grape juice. J. Agric. Food Chem. 35, 519–524.
- Tuckmantel, W., Kozikowski, A.P. and Romanczyk, L.J.Jr. (2008) Studies in polyphenol chemistry and bioactivity. 1. Preparation of building blocks from (+)-catechin. Procyanidin formation. Verma, S. and Singh, S.P. Current and Future Status of Herbal Medicines. Veterinary World, 1, 347-350. https://doi.org/10.5455/vetworld.2008.347-350
- Weir, T.L., Park, S.W. and Vivanco, J.M. (2004) Biochemical and physiological mechanisms mediated by allelochemicals. *Curr. Opin. Plant Biol.* 7(4), 472–479. DOI: 10.1016/j.pbi.2004.05.007
- Weston, L.A. and Duke, S.O. (2003) Weed and crop allelopathy. *Crit. Rev. Plant Sci.* 22 (3-4), 367–389. DOI: 10.1080/713610861
- Yang, F., Yang, Y., Zeng, W. (2020) The Inhibition of Cell Growth Through the EGFR/ERK/MMP-2 Pathway Induced by Ampelopsin in the Human Malignant Melanoma A375 Cell Line. Natural Product Communications. 15(3): 1–9. DOI: 10.1177/1934578X20912864
- Yang, X., Kang, M.C., Lee, K.W., et al. (2011) Antioxidant activity and cell protective effect of loliolide isolated from Sargassum ringgoldianum sub sp. coreanum. Algae. 26(2), 201-208.
- Zaman, F., Iwasaki, A., Suenaga, K. and Kato-Noguchi, H. (2020) Allelopathic potential and identification of two allelopathic substances in *Eleocharis atropurpurea*. *Plant Biosyst.* 155(3): 510-516.
- Zhou, B., Kong, C.H., Li, Y.H., Wang, P. and Xu, X.H. (2013) Crabgrass (Digitaria

sanguinalis) are allelochemicals that interfere with crop growth and the soil microbial community. J. Agric. Food Chem. 61, 5310–5317. DOI: 10.1021/jf401605g

- Zhou, Y., Shu, F., Liang, X., et al. (2014) Ampelopsin induces cell growth inhibition and apoptosis in breast cancer cells through ROS generation and endoplasmic reticulum stress pathway. *PLoS One*. 9(2):89021.
- Zhou, Y., Shu, F., Liang, X., et al. (2014) Ampelopsin induces cell growth inhibition and apoptosis in breast cancer cells through ROS generation and endoplasmic reticulum stress pathway. *PLoS One*. 9(2):89021.

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